



# **The Effects of Extraction Temperature on the Physicochemical Properties of Mangrove-Derived Glucomannan (*Bruguiera gymnorhiza*)**

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**Abstract.** This study investigates the impact of different extraction temperatures on the physicochemical properties of glucomannan derived from mangrove fruits (*Bruguiera gymnorhiza*). Various extraction temperatures ranging from 45°C to 85°C were utilized. Significant differences ( $p < 0.05$ ) were observed in solubility ( $58.41\% \pm 2.45$ ), total reducing sugar content ( $0.39\% \pm 0.09$ ), yield ( $35.13 \pm 2.95$ ), and L\* color value ( $71.97 \pm 1.53$ ), while no significant differences ( $p > 0.05$ ) were found in a\* and b\* color values. These findings have implications for expanding the applications of *Bruguiera* and advancing research on *Bruguiera* glucomannan. Scanning electron microscopy (SEM) analysis revealed an increase in the cross-linking density of glucomannan molecules

**Keywords:** extraction method, physicochemical analysis, food application, mangrove

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## **1. Introduction**

Glucomannan, composed of D-mannose and D-glucose with an  $\alpha$ -1,4-pyranoside bond and acetyl group substitution [1]; [2], is known for its beneficial properties in various sectors, especially in food and pharmaceutical applications [3]; [4]. As a water-soluble non-ionic polysaccharide, glucomannan serves as a natural and renewable polymer dietary fiber [5] and has been widely used as a nutritional

supplement [6].

Several prior investigations have been conducted to extract glucomannan from diverse plant sources such as *Aspergillus oryzae* [7], aloe vera [8]; Salep (*Orchidaceae* family) [1], hyacinth orchid flowers (*Bletilla formosa*) [9]; and Chinese bellflower (*Platycodon grandiflorum* [10]. Additionally, glucomannan is derived from the tubers of several members of the Araceae family, including taro (*Colocasia esculenta*), suweg (*Amorphophallus paeoniifolius*), and sente (*Alocasia macrorrhiza*), salak seeds [11] and Xanthosoma [12]. Different plant sources and extraction techniques yield varied glucomannan characteristics [4]. Glucomannan belongs to the mannan family of polysaccharides, is abundant in nature, particularly in sources like tubers, softwoods (hemicellulose), roots, and various plant bulbs [3].

The plant identified as *Bruguiera gymnorhiza* of mangrove is one of Indonesia's numerous foods categorized as complex carbohydrate sources. Mangrove fruit boasts a high fiber and carbohydrate content [13][14]. The method of glucomannan collection plays a pivotal role since the functional attributes of the final product are linked to process parameters [1]. The heating temperature and pH level have a major impact on the gelation of konjac glucomannan, which is crucial for the use of konjac glucomannan-based products. Glucomannan composites showed a tendency to form gels as the incubation temperature increased from 40°C to 90°C [15]. Exploring the potential development of glucomannan from various plant sources abundant in coastal regions presents an opportunity to investigate effective and efficient extraction methods for enhancing the value of *Bruguiera* glucomannan. This study aimed to investigate a temperature-based extraction method for obtaining glucomannan from *Bruguiera gymnorhiza* and examine its physicochemical properties.

## 2. Methods

### 2.1. *Bruguiera gymnorhiza* Flour

Mangrove fruits utilized in this research were from *Bruguiera gymnorhiza*. The *Bruguiera* mangroves were sourced from the coastal mangrove area in Cilacap, Central Java. These were fruits of the old *Bruguiera* variety measuring around 10-15 cm, displaying a long brownish-green color. Fresh *Bruguiera* fruits were cleaned and peeled. The initial step involved boiling the mangrove *Bruguiera* in water at 80°C for 30 minutes, along with the incorporation of 15% (w/w) hush ash. Subsequently, the mangrove fruit was peeled, and soaked for 48 hours with water changes every 6 hours. The *Bruguiera* mangrove was then cut into 1 - 2 cm thick chips and dried using a cabinet dryer at 70°C for 7 hours. Finally, the dried sliced *Bruguiera* mangroves were ground, and the resulting powder was sieved using an 80-mesh sieve.

### 2.2. Glucomannan Extraction

Extraction of glucomannan from *Bruguiera* flour was performed following the method outlined by [16] with slight modification. *Bruguiera* flour was mixed with distilled water in a ratio of 1:10 (v/w). To optimize glucomannan characteristics, various temperatures ranging from 40 to 90 °C were explored during the extraction process as suggested by previous research [15], The mixing process was conducted at different temperatures, including 45, 55, 65, 75, and 85°C. Subsequently, centrifugation was carried out at 2,800 rpm for 5 minutes. The resulting mixture was treated with 70% technical ethanol at a ratio of 1:5 (v/w) relative to the initial *Bruguiera* mangrove flour weight. The solution was then filtered using filter paper and dried in a drying cabinet at 60°C for 6 hours. The obtained dried *Bruguiera* glucomannan was sieved through an 80-mesh

### 2.3. The Reducing Sugar Measurement

The assessment of reducing sugar content in *Bruguiera* glucomannan was conducted using the Somogyi-Nelsen method, as detailed in the prior study by [17]. Quantifying the reduction of sugar in *Bruguiera* glucomannan was achieved using a spectrophotometer set at a 540 nm wavelength. To determine the reduced sugar content, a standard curve of glucose was established.

### 2.4. The Solubility Measurement

The determination of *Bruguiera* glucomannan solubility followed the methodology outlined by [18], with some modifications introduced into the procedure. The solubility of glucomannan was subsequently assessed.

$$\text{The solubility (\%)} = 1 - \left( \frac{\text{the total sample and the paper} - \text{the initial paper}}{\text{sample initial weight}} \right) \times 100\%$$

### 2.5. The Colour Measurement of *Bruguiera* Glucomannan

The Minolta spectrophotometer method was used to determine the color of *Bruguiera* glucomannan using coordinates L\* (black/white), a\* (red/green), and b\* (yellow/blue) [19]. The device will be calibrated using a whiteboard (standard reference supplied by Konica Minolta) and a standard blackboard before being used. The color characteristics of *Bruguiera* glucomannan (including both control and treatment groups) were evaluated and repeated four times.

### 2.6. The Total Yield Measurement

The *Bruguiera* glucomannan yield was determined according to the method by [4]. The yield is calculated by comparing the initial weight to the final weight of glucomannan multiplied by 100%.

### 2.7. Morphology of Glucomannan SEM

The SEM EXD analysis revealed the fine structure of *Bruguiera* glucomannan [9]. Specifically, the dried *Bruguiera* glucomannan sample was affixed to a copper platform using conductive adhesive and imaged at different magnifications to investigate its morphology.

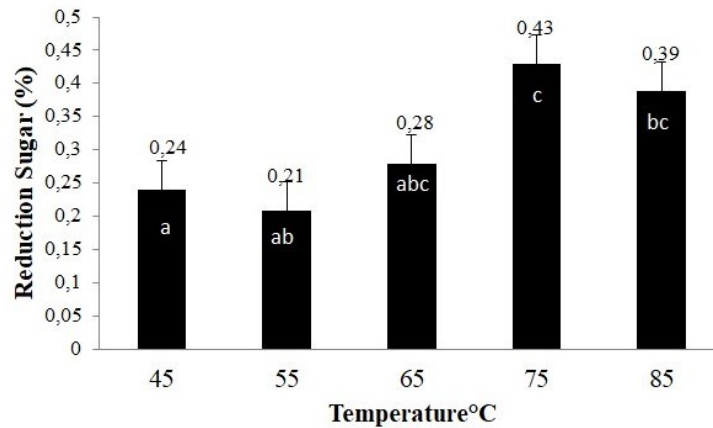
### 2.8. Statistical Analysis

All *Bruguiera* glucomannan measurement results underwent one-way ANOVA analysis using SPSS statistical software. Results were presented as mean  $\pm$  standard deviation (SD). Significance between group means was assessed using Duncan's tests with a statistical significance threshold set at  $p < 0.05$ .

## 3. Results and Discussion

### 3.1. The Reducing Sugar of *Bruguiera* Glucomannan

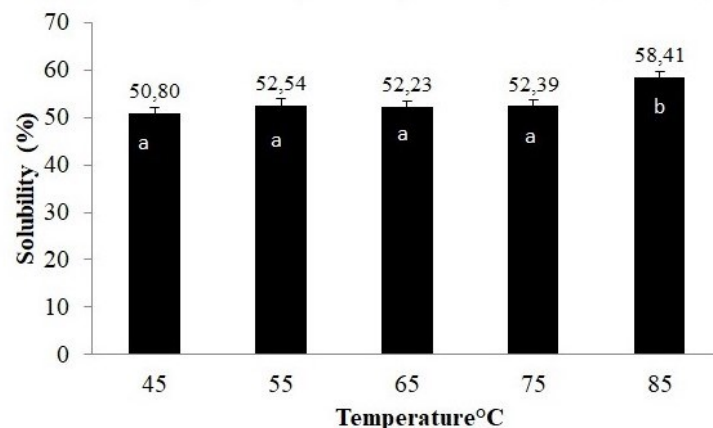
The linear backbone of glucomannan consists of glucose and mannose units, classified as a type of reducing sugar [20]. The reduced sugar of *Bruguiera* glucomannan is depicted in Figure 1. There were significant differences in the total reducing sugar content of *Bruguiera* glucomannan based on extraction temperature ( $p < 0.05$ ). Glucose levels slightly increased with higher temperatures. The total reducing sugar content of *Bruguiera* glucomannan ranged from 0.21 to 0.43%. As stated in [17], extraction temperature influences the hydrolysis process, resulting in changes in the fructose and glucose structure as reducing sugars. In literature [21] glucomannan primarily comprises glucose and mannose units; variations in the ratios of glucose to mannose are attributed to species differences and disparities in processing and treatment methods. The mannose-to-glucose ratio in glucomannan varies depending on the specific source of origin [22].



**Figure 1.** The Result of Reducing Sugar

### 3.2. The Total Solubility of *Bruguiera Glucmannan*

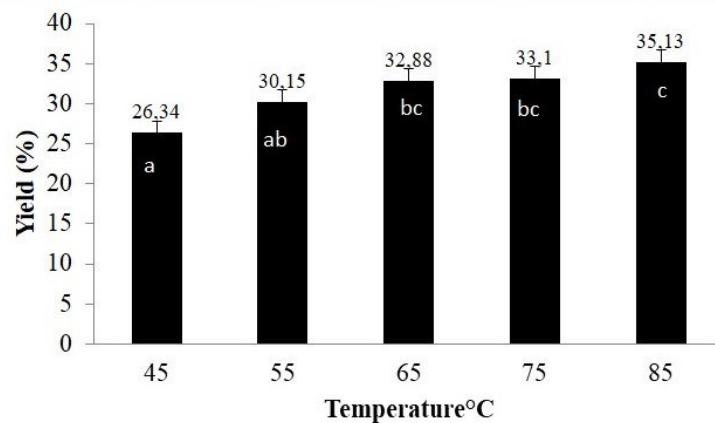
In Figure 2. the solubility of glucomannan from *Bruguiera* was significantly influenced by extraction temperature ( $p < 0.05$ ). The results revealed an overall increase in solubility of *Bruguiera* glucomannan due to variations in extraction temperature. Specifically, an extraction temperature of 85°C demonstrated higher total solubility compared to other treatments. Solubility progressively increased with higher temperatures but no significant difference ( $p > 0.05$ ) was observed among extraction temperatures of 45, 55, 65, and 75°C. These findings suggest that total solubility is impacted by the extraction temperature. As the temperature increases, the solubility of *Bruguiera* glucomannan typically increases, allowing it to dissolve more easily in water. As the temperature rises, the molecular interactions change, leading to greater solubilization and a smoother, more uniform dispersion in the liquid. This relationship between temperature and solubility is crucial in applications where precise control over the consistency of glucomannan-based solutions or gels is required. The higher the solubility of glucomannan, the more convenient it is for use in products. In the literature [16] that the effectiveness of utilizing glucomannan in food relies significantly on its solubility, as it becomes functional upon dissolving in water. According to [3] acetyl groups in glucomannan can hinder intramolecular hydrogen bond formation, thus improving the solubility of glucomannan.



**Figure 2.** The Result of Solubility

### 3.3. The Total Yield of *Bruguiera* Glucomannan

The total yield of *Bruguiera* glucomannan is shown in Figure 3. Significantly higher total yields were observed at extraction temperatures of 65°C and 75°C compared to 45°C and 55°C ( $p < 0.05$ ). The extraction at 85°C yielded the highest overall glucomannan extraction from *Bruguiera*. The yield of glucomannan from *Bruguiera* ranged from 26.34% to 35.17%. Increasing the extraction temperature had a significant impact on the extraction yield ( $p < 0.05$ ). The yield is subject to fluctuations based on the chosen extraction technique. At higher temperatures, the cell walls of plant materials *Bruguiera* containing glucomannan tend to break down more easily, allowing for greater polysaccharide release. This often leads to an increased yield. Yanuriati *et al.* (2017) reported the the total glucomannan yield of fresh porang tubers produced from the direct isolation process ranges from 50 - 65%.



**Figure 3.** The Result of Yield

### 3.4. The Color of *Bruguiera* Glucomannan

The color of *Bruguiera* glucomannan is a crucial factor that plays a significant role in shaping consumer preferences and decisions. The  $L^*$  value color of *Bruguiera* glucomannan is depicted in Figure 4. Different extraction methods yielded varied whiteness degrees of *Bruguiera* glucomannan, as shown in Figure 5, ranging between 71.96 and 74.95. Increasing the extraction temperature significantly impacted the whiteness degree of *Bruguiera* glucomannan ( $p < 0.05$ ), decreasing it due to the Maillard reaction. The redness ( $a^*$ ) color of *Bruguiera* glucomannan is illustrated in Figure 6, indicating the redness of the product. Variation in extraction temperature had no significant effect ( $p > 0.05$ ) on the redness color of *Bruguiera* glucomannan, which displayed values of  $4.04 \pm 0.32$ ,  $4.95 \pm 0.46$ ,  $5.07 \pm 0.48$ ,  $4.93 \pm 0.45$ , and  $5.23 \pm 0.87$  at temperatures of 45, 55, 65, 75, and 85°C, respectively. The yellowness ( $b^*$ ) color of *Bruguiera* glucomannan is also depicted in Figure 6. Changes in extraction temperature did not significantly influence the yellowness color of *Bruguiera* glucomannan, with values of  $22.35 \pm 0.58$ ,  $22.83 \pm 0.72$ ,  $22.37 \pm 2.01$ ,  $21.61 \pm 1.62$ , and  $22.09 \pm 2.09$  at temperatures of 45, 55, 65, 75, and 85°C, respectively. In the literature [19] the coloration of glucomannan can be linked to its heightened solubility and blending capacity, leading to the achievement of a homogeneous texture in composite gels.

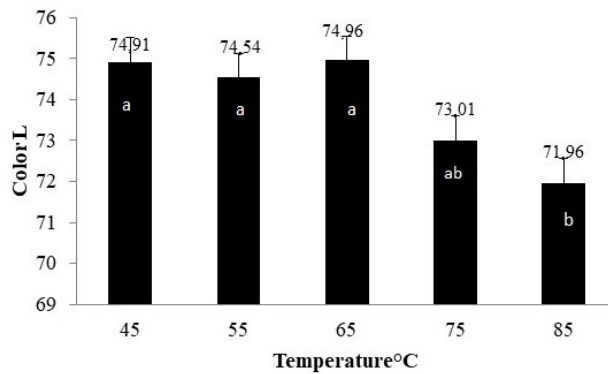


Figure 4. The Color of L\* value

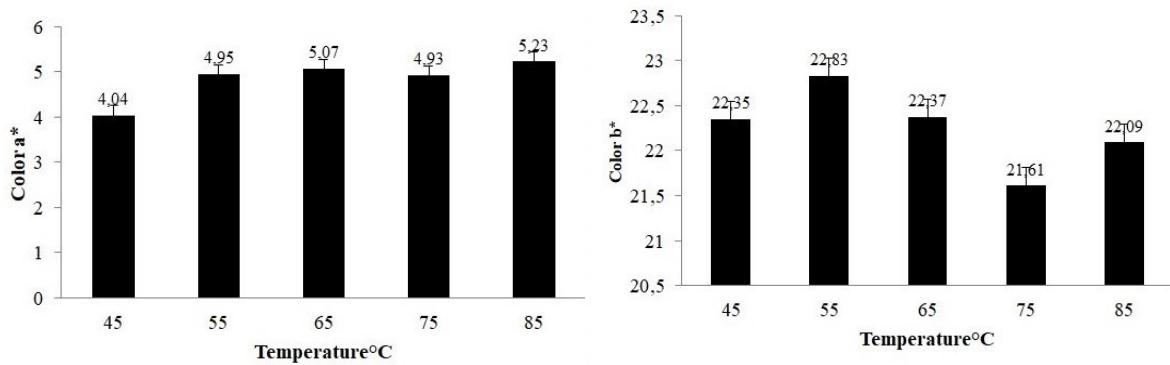


Figure 5. The Redness (a\*) and Yellowness (b\*) Color

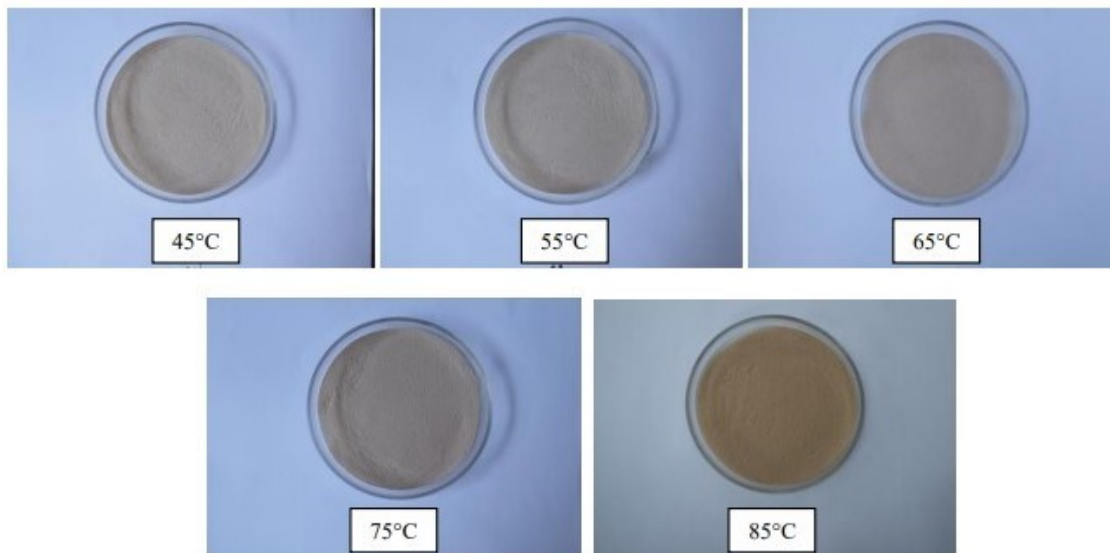


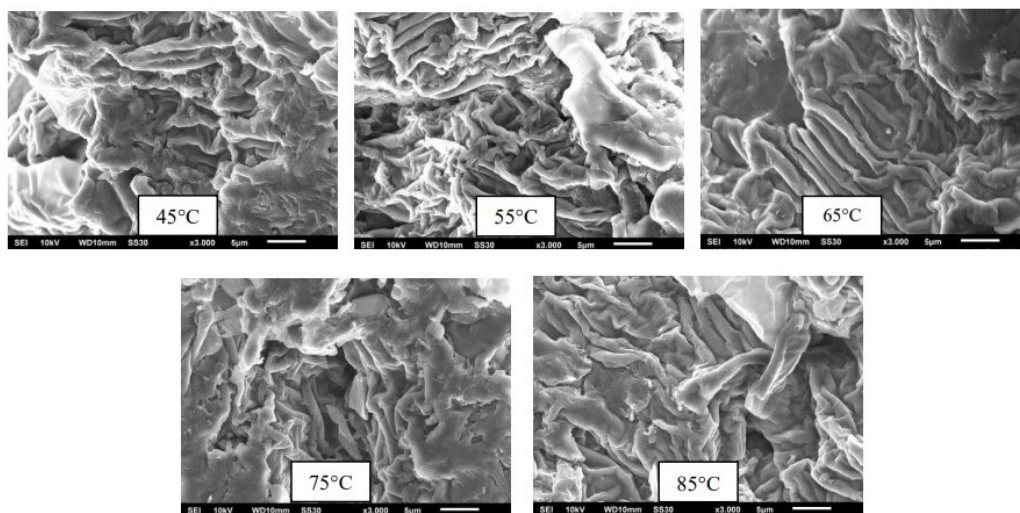
Figure 6. The Color of Glucomannan

### 3.5. The SEM of *Bruguiera* Glucomannan

The differences in surface morphology played a crucial role in shaping the varied functional properties observed in the samples of glucomannan[24]. Figure 7 depicts the structures of glucomannan from *Bruguiera* obtained through varying extraction temperatures. SEM was employed to characterize the morphologies of all *Bruguiera* glucomannan, enabling the observation of particle shapes. Observations of the morphology of *Bruguiera* glucomannan were presented at a magnification of 3000x. The

structure was comprised of a tightly woven network created by glucomannan. The diverse shapes observed in *Bruguiera* glucomannan suggest the onset of the shift from the amorphous to the crystalline phase. [25] reported the transition from the amorphous phase to the crystalline phase in particles derived from glucomannan extracted from fresh porang occurs through the utilization of technical-grade ethanol.

Based on the research findings, there was a significant rise in the cross-linking density of glucomannan molecules at higher extraction temperatures. The microstructure densifies as the extraction temperature increases, aligning with the study [15] that the composites demonstrated a network structure that intertwined as the temperature ranged between 40 and 90°C. The higher incubation temperatures resulted in a relatively elevated cross-linking density of glucomannan molecules [26].



**Figure 7.** The Morphologies of Glucomannan

#### 4. Conclusions

The *Bruguiera* glucomannan content was determined using temperature-based extraction methods. The *Bruguiera* glucomannan was extracted at a temperature of 85°C, resulting in higher levels of total reducing sugar, solubility, and total yield, but lower levels of L\*, a\*, and b\* color values. Varying extraction temperatures can improve the characteristics of *Bruguiera* glucomannan. SEM analysis revealed a significant increase in the density of cross-linking among glucomannan molecules.

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